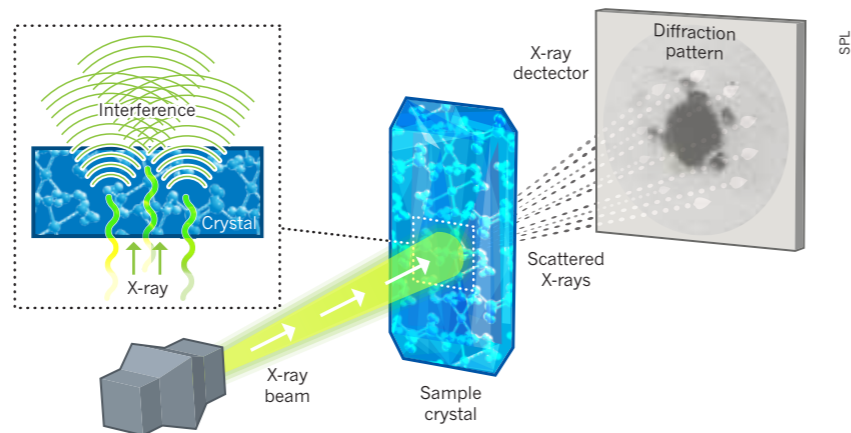


ATOMIC SECRETS

100 YEARS OF CRYSTALLOGRAPHY

BY NICOLA JONES

In 1914, German scientist Max von Laue won the Nobel Prize in Physics for discovering how crystals can diffract X-rays: a phenomenon that led to the science of X-ray crystallography. Since then, researchers have used diffraction to work out the crystalline structures of increasingly complex molecules, from simple minerals to high-tech materials such as graphene and biological structures, including viruses. With improvements in technology, the pace of discovery has accelerated: tens of thousands of new structures are now imaged every year. The resolution of crystallographic images of proteins passed a critical threshold for discriminating single atoms in the 1990s, and newer X-ray sources promise images of challenging proteins that are hard or impossible to grow into large crystals.

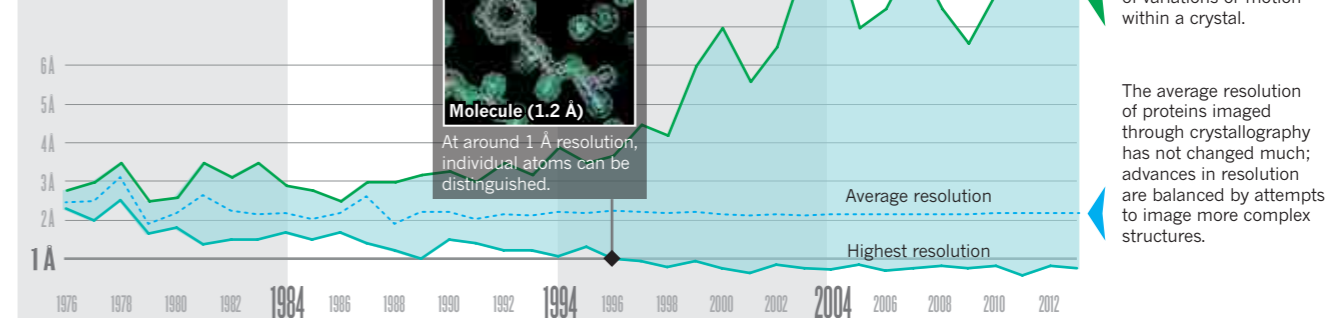


BIRTH OF AN IDEA

Von Laue hit on the idea that when X-rays passed through a crystal, they would scatter off the atoms in the sample and then interfere with each other like waves passing through a breach in a shore wall. In some places, the waves would add to each other; in others, cancel each other out. The resulting diffraction pattern could be used to back-calculate the location of the atoms that scattered the original X-rays. Von Laue and his colleagues proved his theory in 1912 with a sample of copper sulphate.

GETTING CLEARER

Better techniques for both imaging and interpreting data have allowed researchers see finer details in some structures and tackle ever more complicated molecules.



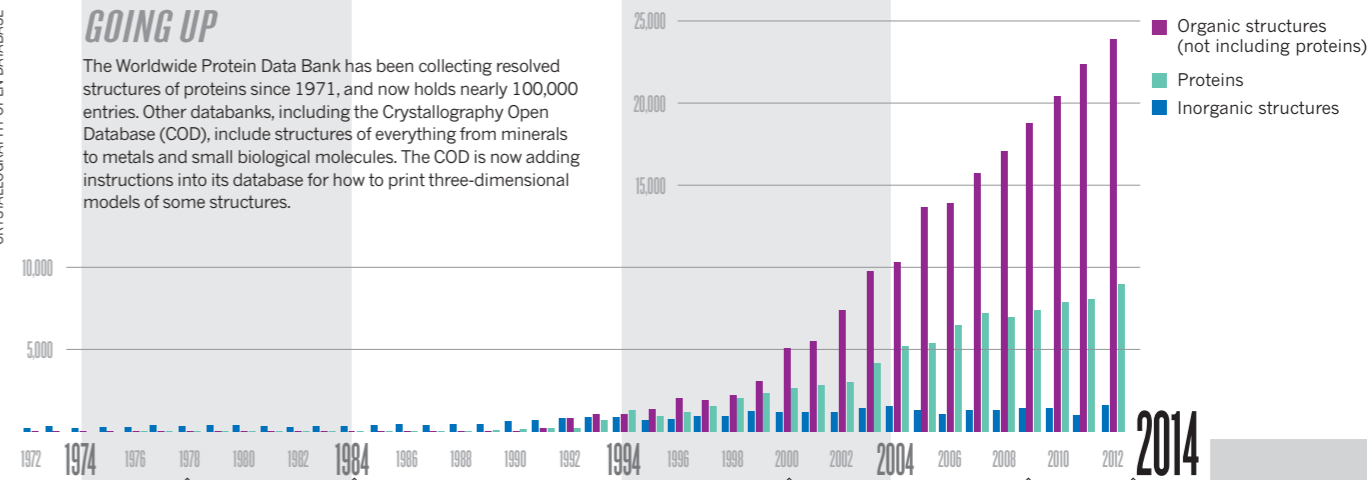
Resolution suffers in images of some complex structures, often because of variations or motion within a crystal.

The average resolution of proteins imaged through crystallography has not changed much; advances in resolution are balanced by attempts to image more complex structures.

GOING UP

The Worldwide Protein Data Bank has been collecting resolved structures of proteins since 1971, and now holds nearly 100,000 entries. Other databanks, including the Crystallography Open Database (COD), include structures of everything from minerals to metals and small biological molecules. The COD is now adding instructions into its database for how to print three-dimensional models of some structures.

SOURCE: WORLDWIDE PROTEIN DATA BANK/CRYSTALLOGRAPHY OPEN DATABASE



1914

1913

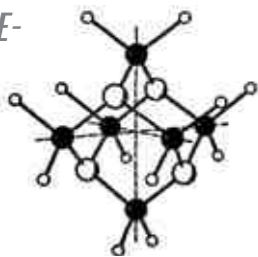
DIAMOND

Diffraction image allowed researchers to confirm the tetrahedral structure of carbon atoms in this famous crystal.

1923

HEXAMETHYLENE-TETRAMINE

The first organic molecule to be imaged, chosen because of its simple cubic symmetry. It proved that molecules, not just atoms, can make up the repeating elements of a crystal.



1925

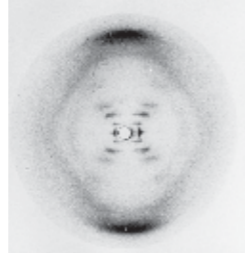
QUARTZ

The determination of the structure of silicate minerals was fundamental to the field of mineralogy.

1952

DNA

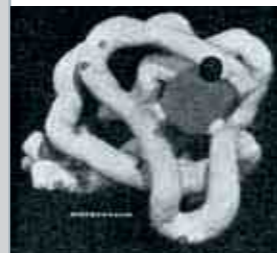
Rosalind Franklin's X-ray image of DNA, known as photo 51, helped James Watson and Francis Crick to create their famous model of the double helix. An atomic-resolution image of the structure proposed in 1953 was not taken until 1980.



1958

MYOGLOBIN

The irregular folds seen in the structure of the first imaged protein were a huge surprise.



1970

SYNCHROTRON

A study of insect muscle at the German Electron Synchrotron (DESY) in Hamburg was the first to use X-rays generated by a synchrotron. The use of these machines caused a boom in crystallography studies.

1965

LYSOZYME

The first enzyme to be imaged, sourced from hen egg whites.



1978

TOMATO BUSHY STUNT VIRUS

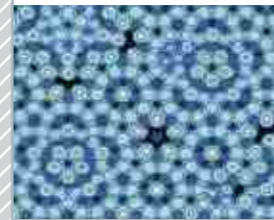
First atomic-scale image of a complete virus: in this case, a plant virus. It revealed structural details that were found to hold true in human pathogens a few years later.



1984

QUASICRYSTALS

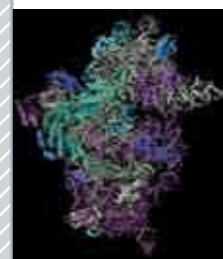
The first crystals were identified with atomic arrangements that do not repeat exactly, defying general wisdom about crystals.



2000

RIBOSOME

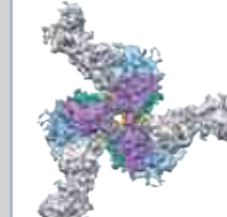
The molecular machine that assembles proteins from instructions encoded in DNA.



2013

HIV TRIMER

An X-ray crystallographic image of the hook that HIV uses to bind to human cells helped to resolve a debate about what this important protein looks like.



2009

X-RAY FREE-ELECTRON LASER

The Linac Coherent Light Source at the SLAC National Accelerator Laboratory in Menlo Park, California, went into operation, opening up a new world of imaging possibilities (see page 604).

THE FUTURE

The 'most wanted' list of proteins that remain to be imaged includes the massive spliceosome, which helps to organize and edit messenger RNA, and the even larger nuclear-pore complex, which serves as a nucleus's gatekeeper.

These structures can contain hundreds of proteins, making them hard to crystallize or keep still for an image.

One strategy is to crystallize bits of these structures and piece them together like a jigsaw; the use of X-ray free-electron lasers should also help.

CRYSTALLOGRAPHY AT 100
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nature.com/crystallography